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INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS GENEVA

# TECHNICAL WORKING PARTY FOR FRUIT CROPS

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WORKING PAPER ON TEST GUIDELINES FOR KIWIFRUIT (Actinidia Lindl.)

Document prepared by experts from New Zealand

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#### I. <u>Subject of these Guidelines</u>

These Test Guidelines apply to all vegetatively propagated fruiting female, polleniser male, hermaphroditic and rootstock varieties of the genus *Actinidia* Lindl.

#### II. Material Required

1. The competent authorities decide when, where and in what quantity and quality the plant material required for testing the variety is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must make sure that all quarantine and customs formalities are complied with. As a minimum., the following quantity of plant material is recommended:

- 8 plants on their own roots or

- 8 plants on a clonal rootstock. The competent authorities to select the most appropriate rootstock.

2. The plant material supplied should be visibly healthy, not lacking in vigor or affected by any important pest or disease. It should preferably not be obtained from *in vitro* culture. If test material is grafted onto a clonal rootstock, there should be information available stating how the rootstock <u>may</u> affect the expression of characteristics. In the case of a female variety the applicant should send in or at least indicate one male variety which flowers at the same time and is compatible with the female variety under test. The male variety should preferably be of the same taxon and at the same ploidy level as the female variety. Hand pollination is suggested to ensure that potential fruit size is achieved.

3. The plant material must not have undergone any treatment unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

#### III. Conduct of Tests

1. To assess distinctness of female and friuting hermaphroditic varieties it is essential that the plants under test bear a satisfactory crop of fruit for at least two growing periods. To assess distinctness of fruit size and fruit shape it is important to ensure adequate seed set, either by hand pollination or by providing sufficient pollinators.

2. To assess distinctness of male and non fruiting varieties it is essential that the plants under test produce two full flowerings over at least two growing periods. If it is claimed that a variety is hermaphroditic, tests should be carried out to determine whether it is self-fertile and self-setting. Pollen viability should be tested separately in addition to flowers being bagged to prevent pollination by outside pollen.

3. The testing should normally be conducted at one place. If any important characteristics of the variety cannot be seen at that place, the variety may be tested at an additional place.

4. The tests should be carried out under conditions ensuring normal growth. As a minimum, each test should include all 8 plants. Separate plots for observation and for measuring can only be used if they have been subject to similar environmental conditions.

5. Additional tests for special purposes may be established.

### IV. Methods and Observations

1. Unless otherwise stated, all observations should be made on 8 plants or 10 parts of 8 plants.

2. For the assessment of uniformity and stability, a population standard of 1% and an acceptance probability of 95% should be applied for varieties resulting from a crossing, and a population standard of 2% with the same acceptance probability for mutations. For a sample size of 8 plants, the maximum number of off-types allowed in both cases would be 1.

3. The shape, size and hairiness of leaves can vary greatly according to the type and vigor of the shoot on which they are borne. Unless specified, the shoots should be replacement canes, i.e., those that will be tied down and retained for the following season's flowering.

4. All observations on the young shoot should be made immediately after flowering, on internodes 10 to 20 cm from the tip of growing shoots.

5. All observations on the stem (including observations on over wintering buds) should be made in the middle third of the replacement stem after leaf fall.

6. All observations on the bud and bud support should be made on dormant canes.

7. All observations on the leaf should be made near the base of the current season's growth on sufficiently mature, but not old leaves. The most basal leaves of a shoot should be excluded since they do not usually attain full size or typical shape.

8. All observations on the presence or absence of red pigment coloration in vegetative organs refer to the general appearance of the organ, irrespective of whether red pigments are present in hairs or in the underlying skin.

9. All observations on the flower should be made on fully opened terminal (king) flowers. The presence of any lateral flowers should be recorded.

10. Unless otherwise stated, all observations on the fruit should be made on fruits at minimum harvest maturity of 6.2°Brix.

V. <u>Grouping of Varieties</u>

1. The collection of varieties to be grown should be divided into groups to facilitate the assessment of distinctness. Characteristics which are suitable for grouping purposes are those which are known from experience not to vary, or to vary only slightly, within a variety. Their various states of expression should be fairly evenly distributed throughout the collection.

2. It is recommended that the competent authorities use the following characteristics for grouping varieties.

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For male varieties:

- (i) Time of beginning of flowering (characteristics 90)
- (ii) Flower: type of coloration (characteristic 53)

For female and hermaphroditic varieties:

- (i) Fruit: size (characteristic 63)
- (ii) Fruit: general shape (characteristic 64)
- (iii) Fruit: skin hairs (characteristic 75)
- (iv) Fruit: outer pericarp color at maturity for consumption (characteristic 83)
- (v) Fruit: time of maturity for harvest (characteristic 91)

#### VI. Characteristics and Symbols

1. To assess distinctness, uniformity and stability, the characteristics and their states, as given in the four UPOV working languages in the Table of Characteristics, should be used.

2. Notes (1 to 9), for the purpose of electronic data processing, are given opposite the states of expression for each characteristics.

3. <u>Legend</u>:

(\*) Characteristics that should be used on all varieties in every growing period over which examinations are made and always be included in the variety descriptions, except when the state of expression of a preceding characteristic or regional environmental conditions renders this impossible.

(+) See Explanations on the Table of Characteristics in Chapter VIII.

# VIII. TABLE OF CHARACTERISTICS

	Characteristic and State	<u>Example</u> <u>Variety</u>	Note
<b>1.</b> (*)	<b>Plant: sex</b> male hermaphrodite	Matua	1 2 2
	remaie	Hayward	3
2.	<b>Plant: fruit setting ( hermap</b> ) absent present	hrodite varieties only)	1 9
3.	Plant: ploidy haploid diploid triploid tetraploid pentaploid hexaploid heptaploid octaploid	Hort16A Hayward	1 2 3 4 5 6 7 8
4.	Plant: vigor very weak weak medium strong very strong	Hayward Matua	1 3 5 7 9
5. (*)	<b>Young shoot: hairs</b> absent present		1 9
<b>6.</b> (*)	Young shoot: density of hair sparse medium dense	<b>s</b> Hayward King	3 5 7
7. (*) (+)	Young shoot: type of hairs downy velutinous tomentose hirsute bristly hispid		1 2 3 4 5 6

<b>8.</b> (*)	Young shoot: red coloration of growing tip absent or very weak weak medium strong very strong	Hort16A King Tomua
9.	Stem: thickness thin medium thick	Hayward Bruno
<b>10.</b> (*)	Stem: color of shoot on sunny side grey white green white grey brown yellow brown light brown red brown purple brown dark brown	King Hort16A Ranger Bruno
11.	<b>Stem: roughness of bark</b> smooth medium rough	Sparkler Meteor
12.	Stem: hairs absent present	
13.	<b>Stem: density of hairs</b> sparse medium dense	
<b>14.</b> (+)	Stem: type of hairs downy velutinous tomentose hirsute bristly hispid	

15. (*)	<b>Stem: size of lenticel</b> small		3
	medium	Hayward	5
	large	Ranger	7
16.	Stem: number of lenticels		
(*)	few	Meteor	3
	medium	Hayward	5
	many	Bruno	7
17.	Stem: color of lenticel		
(*)	greyish white		1
	greyish yellow		2
	greyish brown		3
18.	Stem: size of bud support		
(*)	(bud support diameter in relation	n to stem diameter)	
(+)	small	Sparkler	3
	small to medium	Hayward	4
	medium	King	5
	large		6 7
19.	Stem: position of bud		
(*)	sunken		1
(+)	slightly raised	Ranger	2
	strongly raised		3
<b>20.</b> (*)	Stem: number of hairs visible		
()	few	Havward	3
	medium		5
	many	Hort16A	7
21.	Stem: leaf scar		
(+)	flat	Meteor	1
	shallow	Hayward	2
	deep	Monty	3
22.	Leaf blade: general shape		
(+)	lanceolate		1
	ovate	•• ·	2
	broad ovate	Hayward	3
	very broad ovate	Hort16A	4
	broad obovate	Bruno	5
	very broad obovate	watua	6

23. (*) (+)	Leaf blade: shape of apex emarginate retuse truncate obtuse apiculate acuminate	Bruno Hort16A
24. (*) (+)	Leaf blade: shape of base attenuate acute obtuse rounded truncate cordate	
25. (*) (+)	Leaf blade: arrangement of base (if cordate) far apart slightly apart touching slightly overlapping strongly overlapping	Matua Hort16A Hayward
26.	Leaf blade: density of hairs betwee main veins on upper surface absent or very sparse sparse medium dense very dense	en Hort16A Tomua Bruno Meteor
27.	Leaf blade: density of hairs betwee main veins on lower surface absent or very sparse sparse medium dense very dense	<b>en</b> Hayward Ranger
28.	Leaf blade: profile in cross section concave flat convex	1

29.	Leaf blade: margin entire		1
	denticulate		2
	dentate		3
	serrulate		4
	serrate		5
30.	Leaf blade: puckering/blistering o	n upperside	
(*)	absent or very weak		1
	weak	Hort16A	3
	medium	Hayward	5
	strong		7
	very strong		9
31.	Leaf blade: green color of uppersi	de	
(*)	light		3
	medium	Hayward	5
	dark	Bruno	7
32.	Leaf blade: color of lowerside		
(*)	whitish		1
	light green	_	2
	medium green	Bruno	3
	yellow green	Hayward	4
	yellow brown		5
33.	Leaf blade: glaucosity of lowerside	) )	
(*)	absent		1
	present		9
34.	Leaf blade: variegation		
	absent		1
	present		9
35.	Leaf blade: color of variegation		
	white and green		1
	white, red and green		2
36.	Leaf blade: spines along main		
	vein on lowerside		
	absent		1
	present		9
37.	Leaf : hairs on petiole		-
	absent		1
	present		9

38.	Leaf: density of hairs on petiole sparse		3
	medium	Meteor	5
	dense		7
39.	Leaf: red coloration on upperside of petiole		
	absent or very weak	Hort 16A	1
	weak	Sparkler	3 5
	strong	Tlaywalu	5 7
	very strong		, 9
40.	Flower bud: red coloration of		
	protruding petal ends (at calyx sp	olit)	
	absent or very weak	Hort16A	1
	weak		3
	medium	Hayward	5
	strong	Meteor	0
	very strong		9
41.	Inflorescence: predominant		
(*)	number of flowers	<b>TT</b> 1	1
	one	Hayward	1
	5-5 6-10	Matua Tomuri	2
	> 10	Tomun	4
			-
42.	Pedicel: length		
(*)	very short		1
	short	Matua	3
	long	Hort 16A	כ ד
	verv long	Iomua Iade Moon	9
	very long		,
43.	Pedicel: hairs		
	absent		1
	present		9
44.	Pedicel: length of hairs		_
	short	Hort16A	3
	medium	Hayward	5
	long		/
45.	Flower: number of sepals		
	2 or 3		1
	3-5		2
	> 0		3

<b>46.</b> (*)	Flower: color of sepals white green brown reddish-brown	Hort16A Tomua	1 2 3 4
47.	<b>Flower: sepal hairs</b> absent present		1 9
48.	Flower: length of sepal hairs short medium long		3 5 7
<b>49.</b> (*)	Flower: diameter very small small medium large very large	Matua Hayward	1 3 5 7 9
<b>50.</b> (*) (+)	Flower: arrangement of petals apart touching overlapping	Bruno Hayward	1 2 3
51.	<b>Flower: curvature of petals</b> ( <b>in longitudinal section</b> ) flat curved upwards at tip	Bruno Hayward	1 2
52. (*)	Flower: main color of petals on upper side white greenish white yellowish white yellowish green yellow orange light pink red pink red	Hayward	1 2 3 4 5 6 7 8 9

53. (*)	<ul><li>Flower: type of coloration (upper side)</li><li>single-colored</li></ul>		1
	bicolored	Meteor	2
54. (*)	Flower: shades of color (for single-colored varieties)		
()	absent		1
	present		9
55.	Flower: distribution of color		
(*)	(for single-colored varieties)		
	lighter towards the base		
	ngnier towards the top		Z
56.	Flower: secondary color		
	(for bicolored varieties)		1
	green	Havward	2
	orange		3
	light pink		4
	dark pink	Meteor	5
57.	Flower: distribution of seconda	ary color	
	(for bicolored varieties)		
	marginal	Mataon	1
	bioloned basal spot	Wieteor	2
			5
58.	Flower: color of filament		
	white		1
	light pink		2 3
	dark pink		4
	1		
59.	Flower: color of anther	TT 1	1
	yellow vellow orange	Hayward	1
	grev		$\frac{2}{3}$
	dark purple		4
	black		5
60.	Flower: number of styles		
	few		3
	medium	Hort16A	5
			_

61.	Flower: color of styles white whitish yellow light green		1 2 3
<b>62.</b> (*)	Flower: attitude of styles erect semi-erect both erect and horizontal	Hort16A Hayward	1 2 3
<b>63.</b> (*)	Fruit: size very small small medium large very large	Blake Tomua Hayward Jade Moon	1 3 5 7 9
<b>64.</b> (*) (+)	Fruit: general shape ellipsoidal cylindrical ovoid obovoid globose maliform	Hort16A Bruno Hayward Skelton Katiuscia	1 2 3 4 5 6
<b>65.</b> (*) (+)	<b>Fruit: cross section (at median)</b> circular oblate elliptical	Bruno Hayward	1 2 3
<b>66.</b> (*) (+)	<b>Fruit: general shape of stylar end</b> deeply depressed slightly depressed flat raised protruding	Monty Hayward Bruno Hort16A	1 3 5 7 9
<b>67.</b> (*) (+)	Fruit: shape of shoulder on stalk end square rounded strongly sloping	Hayward Skelton	1 2 3
68.	<b>Fruit: persistance of sepals at ha</b> a absent present	rvest	1 9

69.	Fruit: adherence of skin to flesh weak medium strong		3 5 7
70.	<b>Fruit: lenticels on skin</b> absent present		1 9
71.	Fruit: skin color at harvest matu	rity	
(*)	(fruit still hard)	-	
	light green medium green reddish green greenish brown medium brown reddish brown dark brown	Hort16A Hayward Katiuscia Tomua	1 2 3 4 5 6 7
72.	Fruit: longitudinal stripes or banding on the skin		1
	present		1 9
73. (*)	Fruit: skin color change during ripening absent present		1 9

74. (*)	Fruit: skin color at maturity for consumption		1
	light green		$\frac{1}{2}$
	reddish green		3
	yellow		4
	orange yellow		5
	orange		6
	greenish brown		7
	light brown	Hort16A	8
	raddish brown	Tomus	9
	dark brown	Tomua	10
	purple-red		12
75.	Fruit: skin hairs		
(*)	absent		1
	present		9
76.	Fruit: density of hairs		
(*)	sparse	Hort16A	3
	dense	Hayward	5
	dense	Bruno	/
77.	Fruit: type of hairs		
(*)	downy	Hort16A	1
(+)	velutinous		2
	tomentose	Harmond	3
	hirsute	Hayward Bruno	4
	hispid	Diulio	5
	inspid		0
<b>78.</b>	Fruit: location of hairs		1
(*)	evenly spread		1
	mainly at stylar end		2
79.	Fruit: color of hairs at harvest		1
	white	Hort16A	1
	yellow-brown		23
	medium brown	Havward	3 4
	dark brown	Thuy ward	5
80.	Fruit: adherence of hairs to		
(*)	skin (when rubbed)		
	weak	Hort16A	3
	medium	TT 1	5
	strong	Hayward	7

<b>81.</b> (*)	Fruit: core diameter rela (at largest diameter)	ative to fruit diameter	
()	small	Hort16A	3
	small to medium		4
	medium	Bruno	5
	medium to large	Tomua	6
	large	Hayward	7
82.	Fruit: core shape		
(*)	(in cross section)		
	circular		1
	oblate		2
	elliptic	Hort16A	3
	fluted	Hayward	4
83.	Fruit: outer pericarp col	lor	
(*)	at maturity for consump	tion	1
	light green	Hayward	1
	dark green		2
	medium vellow	Hort16A	З Л
	dark vellow	HOLTOA	+ 5
	vellowish orange		5
	orange		7
	red		8
	red-purple		9
84.	Fruit: inner pericarp col	lor	
(*)	(locules) at maturity for	consumption	
	green	Hayward	1
	greenish yellow		2
	medium yellow	Hort16A	3
	dark yellow		4
	orange-yellow		5
	orange		6
	red		7
	red-purple		8
85.	Fruit: core color at matu	ırity	
(*)	tor consumption		1
	white	TT 1	1
	greenisn white	Hayward	2
	yellow-white	Hort16A	3
	orange		4
	red-purple		5

<b>86.</b> (*)	Fruit: sweetness (Brix level) at maturity for consumption very low low medium high very high	Jade Moon Hayward Tomua Hort16A	1 3 5 7 9
87.	Fruit: titratable acidity (as citric a at maturity for consumption low (< 0.6%) medium (0.6-1.0%) high (> 1.0%)	acid)	3 5 7
88.	Fruit: vitamin C content low low to medium medium medium to high high	Hayward Bruno	3 4 5 6 7
<b>89.</b> (*)	<b>Time of vegetative budbreak</b> early medium late	Tomua Hayward	3 5 7
<b>90.</b> (*)	<b>Time of beginning of flowering</b> early medium late	Hort16A Bruno Hayward	3 5 7
91.	Time of maturity for harvest		
(*)	very early early medium late very late	Tomua Bruno Hayward	1 3 5 7 9

# IX. Explanations to the Table of Characteristics

### X. Literature

Astridge, S.J. 1975: Cultivars of Chinese gooseberry (*Actinidia chinensis*) in New Zealand. Economic Botany 29: 357-360.

Bellini, E.; F. Monastra, 1986: Propagazione, problemi vivaistici, scelta varietale e miglioramento genetico dell'actinidia. pp. 43-83. *In*: G. Bargioni, F. Lalatta and A. Febi (coord.). *Incontro frutticolo la coltura dell'actinidia*. Atti del Convegno, Verona, 29 Aprile 1986. Verona, Cassa di Risparmio di Verona, Vicennza e Belluno per l'Agricoltura.

Bergamini, A.; F. Monastra 1989: Schede per lo studio dell'actinidia in uso presso l'Istituto sperimentale per la Frutticoltura di Roma. *Annali dell'Istituto Sperimentale per la Frutticoltura* 20: 121-134.

Cui, Z.-X. 1993: [*Actinidia* in China] (in Chinese) Jinan, China: Shandong Scientific and Technology Press.

Ferguson, R (draft) for inclusion in Handbook of Fruit Cultivars, ASHS

Japanese National Test Guidelines for Kiwifruit, Matatabi and Kokuwa 1995

Testolin, R.; V. Crivello 1987: *Il kiwi e il suo mondo*. Venezia: federazione Regionale Coltivatore Diretti del Veneto; Centrol Regionale IRIPSA-Quadrifoglio.

Union Internationale pour la Protection des Obtentions Vegetales 1985. Guidelines for the conduct of tests for distinctness, homogeneity and stability. Kiwifruit, Actinidia, kiwi (*Actinidia chinensis* Pl.) TG/98/3.

Valmori, I. 1991: Nuove varieta in frutticoltura. Bologna: Edizioni Agrficole.

Zhang, J.; T.G. Thorp 1986: Morphology of nine pistillate and three staminate New Zealand clones of kiwifruit (*Actinidia deliciosa* (A. Chev.) C.F. Liang et A.R. Ferguson var. *deliciosa*). New Zealand Journal of Botany 24: 589-613.

# XI. <u>Technical Questionnaire</u>

			Reference Number (not to be filled in by the applicant)
	to be completed in	TECHNICAL QUESTION connection with an applicati	NAIRE on for plant breeders' rights
1.	Species	Actinidia Lindl.	
		KIWIFRUIT	
2.	Applicant (Name and a	ddress)	
3.	Proposed denomination	or breeder's reference	

4.	Information on origin, maintenance and reproduction of the variety						
4.1	Bree	Breeding method					
	(a)	a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?					
		Yes [] No []					
	(b)	Has such author	zation been obta	ined?			
		Yes []	No	[]			
	If th	e answer to that q	uestion is yes, ple	ease attach a co	opy of such an	authorization.	
4 2	Orio	n					
1.2	(a)	Seedling of unk	nown parentage			[]	
	<ul> <li>(a) Secting of unknown parentage</li> <li>(b) Produced by controlled pollination</li> <li>(indicate parent varieties)</li> </ul>					[]	
		<ul> <li>Seed bearing parent</li> </ul>					
	– Pollen parent []					[]	
	(c) Produced by open pollination of [] (indicate seed bearing parent plant)			[]			
	(d) Mutation or sport from (indicate parent variety)						
	[]				[]		
	(e)	(e) Discovery (indicate where and when)					
						[]	

4.3.	In vitro propagation		
	The plant material has been obtained by in vitro propagation	yes no	[]
4.4	Virus status		
	The variety is		
	(a) virus free (indicate viruses)		[]
			••••
	(b) virus tested (indicate against which virus)		
	(c) The virus status is unknown		
4.5	Pollinator [for female varieties]		
	Good pollinators are the following varieties:		
4.6	Other information		

Characteristics of the variety to be indicated (the number in brackets refers to the 5. corresponding characteristic in Test Guidelines; please mark the state of expression which best corresponds). **Example Varieties** Characteristics For male varieties 5.1 Flower: type of coloration (upper side) (53) single-colored 1[] bicolored Meteor 2[] 5.2 Time of beginning of flowering (90) Hort16A early 3[] medium Bruno 5[] late Hayward 7[] For female and hermaphrodite varieties 5.3 (63) Fruit: size 1[] very small Blake small 3[] medium Tomua 5[] large Hayward 7[] Jade Moon very large 9[] 5.4 Fruit: general shape (64) ellipsoidal Hort16A 1[] cylindrical Bruno 2[] ovoid Hayward 3[] obovoid Skelton 4[] globose Katiuscia 5[] maliform 6[]

	Characteristics	Example Varieties		
5.5 (75)	Fruits: skin hairs			
	absent		1[	]
	present		9[	]
5.0	Fruite automonican calculation			
5.0 (83)	Fruit: outer pericarp color at maturity for consumption			
	light green	Hayward	1[	]
	dark green		2[	]
	greenish yellow		3[	]
	medium yellow	Hort16A	4[	]
	dark yellow		5[	]
	yellowish orange		6[	]
	orange		7[	]
	red		8[	]
	red-purple		9[	]
5.7 (91)	Time of maturity for harvest			
	very early		1[	]
	early		3[	]
	medium	Tomua	5[	]
	late	Bruno	7[	]
	very late	Hayward	9[	]

6. Similar varieties and differences from these varieties					
D	enomination of similar variety	Characteristic in which the similar variety is different <sup>o)</sup>	State of expression of similar variety	State of expression of candidate variety	
0)	In the case of identifier the difference.	ntical states of expression	ns of both varieties, plea	se indicate the size of	
7.	Additional inform	ation which may help to	distinguish the variety		
7.1	Resistance to pest	s and diseases			
7.2	Special conditions	for the examination of	the variety		
7.3	Other information				
A representative color photo of the variety should be included in the Technical Questionnaire.					

[End of document]